

# Informal Examiner Amdt. Scan AS SPEC

mercaptoethanol, pH 8.0) and is kept at 37°C after addition of 180 units of ExoIII nuclease. Ten µl portions of the solution are sampled every 30 seconds and transferred to tubes containing 10 µl of MB buffer (40 mM NaCl, 2 mM ZnCl<sub>2</sub>, 10% glycerol, pH 4.5) in an ice bath. After sampling, the 10 tubes are kept at 65°C for 10 minutes to inactivate the enzyme. Then, 5 units of mung bean nuclease is added and kept at 37°C for 30 minutes. From one original plasmid, ten different kind of DNA fragments are recovered by agarose gel electrophoresis. The degree of deletion of each fragment varies. The terminals of the recovered DNAs are smoothed with Klenow enzyme to subject to ligation reaction at 16°C overnight, and by using resulting DNA, E. coli DH5α is transformed to obtain clones. The plasmids are prepared from the various clones obtained, and nucleotide sequences are determined by using luminescence primer cycle sequence kit (Applied Biosystems corp.) with an automatic sequencer.

As a result, it was found that the nucleotide sequence of the cDNA in pRH1 derived from Phaffia rhodozyma consists of 1,099 base pairs (~~SEQUENCE ID No. 4~~)<sup>SEQ ID NO: 4</sup>, and there is an open reading frame which encodes a polypeptide having 251 amino acids (which corresponds the region from A to B in Figures 4 and 5). It was also found that the nucleotide sequence of the cDNA in pHP1 derived from Haematococcus pluvialis consists of 1,074 base pairs (~~SEQUENCE ID No. 5~~)<sup>SEQ ID NO: 5</sup>, and there is an open reading frame which encodes a polypeptide having 259 amino acids (which corresponds the region from C to D in Figures 6 and 7). The amino acid sequences expected from these open reading frames are investigated by analyzing homology in the Gene Bank. Both of the amino acid sequences of Ph. rhodozyma and Ha. pluvialis have

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(EXAMPLE 9) Isolation of the IPP isomerase gene of  
Saccharomyces cerevisiae by PCR method

Based on the nucleotide sequence of the IPP isomerase gene of  
S. cerevisiae reported in the aforementioned reference (Anderson,  
M. S., Muehlbacher, M., Street, I.P., Profitt, J., Poulter, C.  
D., "Isopentenyl diphosphate: dimethylallyl diphosphate isomerase  
- an improved purification of the enzyme and isolation of the  
gene from Saccharomyces cerevisiae", J. Biol. Chem., 264:19169-  
19175(1989)), the primers below were synthesized. (SEQ ID NOS 7 and 8, respectively)

Primer No. 1 5'-TCGATGGGGGTTGCCTTTCTTTTTCGG-3'

Primer No. 2 5'-CGCGTTGTTATAGCATTCTATGAATTGCCC-3'

The procedure was designed to obtain PCR amplified IPP  
isomerase gene having TaqI sites at the upstream terminal and  
AccII region at the downstream terminal. Thirty cycles of PCR is  
performed with 200 ng of total DNA of S. cerevisiae and PfuDNA  
polymerase (STRATAGENE). To express the IPP isomerase gene  
obtained by PCR in E. coli, it is digested with both TaqI and  
AccII. Then, the gene was inserted into ClaI sites and SmaI  
sites of pBluescript KS+ vector. The resulted plasmid was named  
pSI1 (Figure 11). This DNA derived from S. cerevisiae had a  
nucleotide sequence consisting of 1,058 bp (SEQ ID NO: 6)  
(~~SEQUENCE ID NO. 6~~),  
and contained a gene which encodes IPP isomerase consisting of  
288 amino acids (SEQ ID NOS: 4, 5, and 3)  
(corresponds from E to F in Figures 8 and 9).

(EXAMPLE 10) Increase of lycopene production amount by  
introducing the IPP isomerase gene

Into the lycopene-producing E. coli JM101 strain (abbreviated  
as L hereafter) which contains pACCRT-EIB (Figure 10), pSPORT1